



Online Version ISSN: 1314-412X
Volume 3, Number 3
September 2011

AGRICULTURAL SCIENCE AND TECHNOLOGY

2011

An International Journal Published by Faculty of Agriculture,
Trakia University, Stara Zagora, Bulgaria

Editor-in-Chief

Tsanko Yablanski
Faculty of Agriculture
Trakia University, Stara Zagora
Bulgaria

Co-Editor-in-Chief

Radoslav Slavov
Faculty of Agriculture
Trakia University, Stara Zagora
Bulgaria

Editors and Sections

Genetics and Breeding

Atanas Atanassov (Bulgaria)
Ihsan Soysal (Turkey)
Max Rothschild (USA)
Stoitcho Metodiev (Bulgaria)

Nutrition and Physiology

Nikolai Todorov (Bulgaria)
Peter Surai (UK)
Zervas Georgios (Greece)

Production Systems

Dimitar Pavlov (Bulgaria)
Dimitar Panaiotov (Bulgaria)
Jordan Staikov (Bulgaria)
Georgi Zhelyazkov (Bulgaria)

Agriculture and Environment

Georgi Petkov (Bulgaria)
Ramesh Kanwar (USA)

Product Quality and Safety

Marin Kabakchiev (Bulgaria)
Stefan Denev (Bulgaria)

English Editor

Yanka Ivanova (Bulgaria)

Scope and policy of the journal

Agricultural Science and Technology /AST/ – an International Scientific Journal of Agricultural and Technology Sciences is published in English in one volume of 4 issues per year, as a printed journal and in electronic form. The policy of the journal is to publish original papers, reviews and short communications covering the aspects of agriculture related with life sciences and modern technologies. It will offer opportunities to address the global needs relating to food and environment, health, exploit the technology to provide innovative products and sustainable development. Papers will be considered in aspects of both fundamental and applied science in the areas of Genetics and Breeding, Nutrition and Physiology, Production Systems, Agriculture and Environment and Product Quality and Safety. Other categories closely related to the above topics could be considered by the editors. The detailed information of the journal is available at the website. Proceedings of scientific meetings and conference reports will be considered for special issues.

Submission of Manuscripts

All manuscript written in English should be submitted as MS-Word file attachments via e-mail to ascitech@uni-sz.bg. Manuscripts must be prepared strictly in accordance with the detailed instructions for authors at the website <http://www.uni-sz.bg/ascitech/index.html> and the instructions on the last page of the journal. For each manuscript the signatures of all authors are needed confirming their consent to publish it and to nominate an author for correspondence. They have to be presented by a submission letter signed by all authors. The form of the submission letter is available upon request from the Technical Assistance or could be downloaded from the website of the journal. All manuscripts are subject to editorial review and the editors reserve the right to improve style and return the paper for rewriting to the authors, if necessary. The editorial board reserves rights to reject manuscripts based on priorities and space availability in the journal.

Subscriptions

Agricultural Science and Technology is published four times a year. The subscription price for institutions is 80 € and for personal subscription 30 € which

include electronic access and delivery. Subscription run for full calendar year. Orders, which must be accompanied by payment may be sent direct to the publisher:

Trakia University
Faculty of Agriculture, Bank account:
UniCredit Bulbank,
Sofia BIC: UNCRBGSF

IBAN: BG29UNCR76303100117681
With UniCredit Bulbank Stara Zagora

Internet Access

This journal is included in the Trakia University Journals online Service which can be found at www.uni-sz.bg.

Copyright

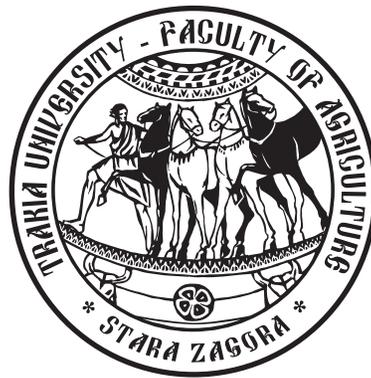
All rights reserved. No part of this publications may be translated into other languages, reproduced or utilized in any form or by any means, electronic or mechanical, including photocopying or any information storage and retrieval system without permission in writing from the publisher.

Address of Editorial office:

Agricultural Science and Technology
Faculty of Agriculture, Trakia University
Student's campus, 6000 Stara Zagora
Bulgaria
Telephone.: +359 42 699330
+359 42 699446
<http://www.uni-sz.bg/ascitech/index.html>

Technical Assistance:

Nely Tzvetanova
Telephone.: +359 42 699446
E-mail: ascitech@uni-sz.bg



*AGRICULTURAL
SCIENCE AND TECHNOLOGY*

2011

An International Journal Published by Faculty of Agriculture,
Trakia University, Stara Zagora, Bulgaria

Development of the caecal microbiota in rabbits weaned at different age

B. Bivolarski¹, G. Beev², S. Denev^{2*}, E. Vachkova¹, G. Kostadinova³, T. Slavov⁴

¹Department of Pharmacology, Animal Physiology & Physiological Chemistry, Faculty of Veterinary Medicine, Trakia University, 6000 Stara Zagora, Bulgaria

²Department of Biochemistry & Microbiology, Faculty of Agriculture, Trakia University, 6000 Stara Zagora, Bulgaria

³Department of Applied Ecology and Animal Hygiene, Faculty of Agriculture, Trakia University, 6000 Stara Zagora, Bulgaria

⁴Department of Morphology, Physiology and Animal Nutrition, Faculty of Agriculture, Trakia University, 6000 Stara Zagora, Bulgaria

Abstract. The experiment was conducted to study the post-natal development of the caecal microbiota in rabbits weaned at different age. A total of 60 healthy New Zealand White rabbits of both sexes, born the same day, were used in the experiment (after controlling for the effect of litter origin and weaning weight and variability). Rabbits were weaned both at 21 days (W21 group, 30 litters) and at 35 days (W35 group, 30 litters) of age. The weaned animals were randomly housed in wire net cages measuring in well-controlled experimental facility. They received standard commercial pelleted diet without antibiotics. Feed and drinking water were available *ad libitum*. Results of the microbiological examination of the caecal contents indicated that rabbits weaned at 35 day had higher total bacterial count (TBC) per g of caecal content, in comparison with rabbits weaned at 21 day ($P < 0.001$). The TBC in the caecum of earlier and later weaned rabbits after weaning increased significantly ($P < 0.001$). The obligate anaerobic bacteria, particularly *Bacteroides* spp. constitute an important group of microorganisms in the rabbit caecum. The population of *Bacteroides* spp. increased with advancing of age. The differences between groups on days 35, 42 and 49 were statistically significant ($P < 0.001$). Sporulating bacteria and especially *Cl. perfringens* was present in low variable amounts in all the caecal samples obtained from healthy animals. Caecal counts of *Cl. perfringens* at weaning (21 and 35 day) were very low (1.656 and 1.654 log₁₀ CFU/g, respectively) and not affected by weaning age. To the end of the study, earlier weaned rabbits had higher caecal count of *Cl. perfringens* ($P < 0.01$). *Enterococcus* spp. and coliforms, including *E. coli* are an important part of the caecal microbial population of rabbits. The caecal number of coliforms was considerably high at weaning, then decreased linearly and stabilized on low level at day 49. Our study demonstrated the absence of *Lactobacillus* spp. in the rabbit caecal tract. The pH of the caecal content fell linearly throughout the experiment - there are not significant differences between groups at days 21 and 49. Compared to the W21 group, rabbits in the W35 group, had a higher live body weight ($P < 0.001$) and low mortality during the trial.

Keywords: rabbits, caecal microbiota, weaning age

Abbreviations: CE – caecal ecosystem, GIT – gastrointestinal tract, CFU – colony forming units, TBC – total bacterial count, CP – crude protein, BW – body weight, CW – weight of the caecal wall, CC – caecal content

Introduction

The rabbit caecal ecosystem (CE) plays a key role in the digestive physiology, both for health and digestive efficiency, because of its size (40% of the whole tract content) and of the highly active microbiota (Gidenne, 2003; Gidenne et al., 2004; Monteils et al., 2008). In this ecosystem there are complex interactions among the biotope (i.e. the caecum) and the biocenosis (the microbiota), and its functioning still remains poorly known (Gidenne et al., 2008).

The caecum is very large, compared with the rest of the gut (Stevens and Hume, 1995) and forms a spiral that fills the abdominal cavity. Previous studies on rabbit caecal microbiota (CM) revealed that bacterial species are mainly strictly anaerobic, but they were mostly performed using classical culture-based techniques (Monteils et al., 2008). The microbial composition of rabbit CE is changed with advancing of age, particularly in association with the weaning period.

Weaning is a stressful period for rabbits when healthy caecal microflora is not yet established (Harcourt-Brown, 2002). The period around weaning is a very critical time for young rabbits as they are highly sensitive to multifactor digestive disorders (Kovacs et al., 2008). Up to 18-20 days, rabbits ingest only milk, while from this point onwards they begin to consume solid feed as well, initially in

small quantities and subsequently, as their mother's milk yield diminishes in ever increasing quantities (Nizza et al., 2002). During this period, the gastrointestinal tract (GIT) of rabbits is colonized by an abundant microflora, predominantly by strictly anaerobic bacteria (Marounek et al. 2000a). The fermentative activity of caecum begins to develop (Padilha et al., 1995, 1996; Piattoni et al., 1995) and the enzymatic digestive activities show important changes (Marounek et al., 1995). The microbial and fermentative activities in the CE of young rabbits play a key role in the prevention of digestive disorders (Carabaño et al., 2006). The pre-weaning milk/feed ratio affects the microbiological and biochemical characteristics of the caecal content and also post weaning performance of rabbits (Nizza et al., 2001). On the other hand, an earlier intake of solid feed could accelerate the maturation of digestive enzymes and also reduce the risk of digestive disorders (Maertens and De Groote, 1990; Scapinello et al., 1999).

In commercial rabbit rearing, weaning of litters usually occurs from 30 to 35 days of age (Xiccato et al. 2003). An earlier weaning of young rabbits has some advantages for both kits and dams: reduces pathogen transmission by limiting contacts between litters and does and incidence of digestive disorders (Schlout, 1988; Gidenne and Fortun-Lamothe, 2002) specific starter diets could better cover kit nutritional requirements (Gutiérrez et al., 2002) improves the body

* e-mail: stefandenev@hotmail.com

condition of the doe and its health status (Pascual, 2001; Xiccato et al., 2000, 2001; Gidenne and Fortun-Lamothe, 2002) and shorter lactation periods could reduce the doe energy output for milk production and the consequent body energy deficit (Parigi-Bini and Xiccato, 1998). On the other hand, early weaning could impair the welfare of dams and litters as well as the viability of kits that are too young, even though in wild rabbits litters may be completely weaned at 23 or 24 days of age because the does, often mated soon after kindling, are going to have their successive kindling (Hudson et al., 2000).

Analysis of the literature shows that during the last decade, much research has been conducted to study the development of digestive activity around weaning (Dojana et al., 1998; Debray et al., 2001a, 2001b; Pinheiro et al., 2001; Gidenne and Fortun-Lamothe, 2002; Gidenne et al., 2004); effects of different nutritional strategies on caecal fermentation (Marounek et al., 2000b; Xiccato et al., 2003; Lavrencic, 2007); and post weaning health and performance (Xiccato et al., 2000; Hampson et al., 2001; Gidenne and Licois, 2005; Feugier et al., 2006; Alvarez et al., 2007; Gomez-Conde et al., 2007; Carabano et al., 2008). A role of caecal microbial activity has been hypothesized but not studied in details (Bennegadi-Laurent et al., 2004). Little is still known about the effects of different biotic and abiotic factors, including weaning, on the caecal microbial ecosystem (Kovacs et al., 2002, 2004, 2006, 2008).

The objective of the current experiment was to study the post-natal development of caecal microbiota in rabbits weaned at different age.

Material and methods

Animals, Housing and Diet

The experiment was performed at the Section of Animal Physiology, Faculty of Veterinary Medicine, Trakia University, Stara Zagora, Bulgaria, between March and May 2009. The rabbits were taken from a commercial farm of the Agricultural Research Institute, located around Stara Zagora. New Zealand White does and their progeny were used in the experiment. All animals were healthy, i.e. no diarrhoea or losses of live weight were observed. The animals were kept in standard wire net cages for reproducing does. At one end of the cage there was a nest box, with a creep-hole that could be closed. The nest box contained a nest tray. The does were fed a standard medication-free pelleted diet. Feed and drinking water were available *ad libitum*. From 14 day of age, the young rabbits had free access to their mothers feed. The temperature of the room was 20-22°C, humidity about 65%, and the daily lighting period was 14 h.

A total of 60 healthy New Zealand White rabbits of both sexes, born the same day at the commercial farm, were used in the experiment (after controlling for the effect of litter origin and weaning weight and variability). Young rabbits were weaned either at 21 days (W21 group, n=30) or at 35 days (W35 group, n=30) of age. The weaned animals were randomly housed in standard cages (800 x 600 x 400 mm) (5 animals per cage) in a well-controlled experimental facility. All cages were equipped with feeding hoppers and drinking nipples. Before the experiment the facility was cleaned and disinfected. The ambient temperature was maintained between 20 and 22°C and relative humidity about 65%. The daily lighting period was 14 h. The weaned rabbits from both groups had access to the same standard commercial pelleted diet, without antibiotics. The ingredients and chemical composition of the diet are listed in Table 1. Throughout the experimental period feed and water were provided

Table 1. Ingredients and chemical composition of the diet

Ingredients	%
Alfalfa meal	20.00
Corn	5.00
Wheat	5.00
Barley	23.00
Oats	4.00
Soybean meal (44% CP)	3.50
Extracted sunflower meal	16.00
Wheat bran	19.00
Sugar-beet pulp	2.00
Vitamin-mineral premix*	0.50
Limestone	1.00
Dicalcium phosphate	0.14
Sodium chloride	0.30
Acidifier (Biotronic®SE)	0.25
Dry binder (Kembind®)	0.30
Antioxidant (Endox®)	0.01
Chemical composition**	
Dry mater,%	88.00
Digestible energy, kcal/kg	2 590
Crude Protein,%	18.00
Crude Fat,%	2.70
Crude fibre,%	13.00
Ash,%	5.00
Lysine,%	0.77
Methionine + Cystine	0.65
Ca	1.00
P	0.50

* Vitamine-mineral premix per kg: Vitamins: A 10 000 IU; D₃ 1 000 IU; E 50 mg; B₁ 5 mg; B₂ 14 mg; D-panthotenic acid 25 mg; B₆ 10 mg; B₁₂ 0.04 mg; PP 80 mg; H 0.3 mg; Folic acid 2 mg; K 8 mg; Choline 1 300 mg; Minerals: Fe 160 mg; I 2.5 mg; Co 0,6 mg; Cu 40 mg; Mn 120 mg; Zn 120 mg; Se 0,2 mg.

** AOAC (2000)

without restriction (*ad libitum*). The health status of the young rabbits before and after weaning was strictly controlled, both to ensure animal welfare and to avoid bias in the results. Mortality was checked daily, while morbidity was assessed weekly through an individual control of all clinical signs of digestive troubles, such as transitory diarrhoea, presence of mucus in excreta, abnormal behaviour, etc., over the whole period of experiment. No signs of disease were registered during the experimental period. All care and experimental procedures involving animals followed the guidelines defined in the Guide to the Care and Use of Laboratory Animals (1996) and the trials were agreed by the Animal Use and Care Advisory Committee of the Faculty of Veterinary Medicine, Trakia University, Bulgaria.

Sampling procedures

Five healthy rabbits, selected randomly from each group, were examined: Group A on days 21, 28, 35, 42, 49 and Group B on 35, 42 and 49 after birth. The feed was removed at 19.00 h on the day before sampling, but water was still available *ad libitum*. The animals were euthanized (between 9:00 and 10:00 a.m.) by an overdose of carbon dioxide and bled. After opening the abdominal cavity the digestive tract was removed and the caecum was separated by tying

off the two extremities with nylon string to prevent movement of the digesta and immediately transported to the laboratory of microbiology in a pre-warmed (39°C) container. The caecum was weighted before and after emptying to determine the weight of its content. The samples were taken from the caecal content with sterile precautions and subjected to microbiological analysis. The manipulations were done under the constant flushing of the oxygen free CO₂ gas to assure anaerobic conditions and the temperature was kept at 39°C.

Microbiological analyses

Immediately after sampling serial 10-fold dilutions were prepared with sterile pre-reduced anaerobic dilution solution, containing (g/l): (K₂HPO₄ × 3H₂O - 4,105; KH₂PO₄ - 1.636; Cysteine hydrochloride - 0.5; Gelatin - 2.0; Resasurin - 1.0), according to procedure described by Holdeman and Moore (1977). The serially diluted caecal content were used for determination of the total bacterial count (TBC); anaerobic (*Bacteroides*, *Clostridium*, *Lactobacillus* spp.); facultative anaerobic organisms (*Enterococcus* spp., and Coliform counts). The different dilutions (10⁸⁻⁹⁻¹⁰ for TBC; 10⁶⁻⁷⁻⁸ for bacteroides; 10³⁻⁴⁻⁵ for enterococci; 10²⁻³⁻⁴ for clostridia, lactobacilli and coliforms) were smeared onto the surface of the selective culture media, previously reduced by placing under anaerobic condition (Gas Pak[®] anaerobic system) for 18 to 24 h prior to use. The obligate anaerobe *Bacteroides* spp. were cultured on Schaedler's Agar (BBL[®], USA), the selectivity of which was increased by the addition of esculin, neomycin and Fe-ammonium citrate (Merck, Germany) (Kovacs et al., 2004, 2006); *Lactobacillus* spp. were cultured on MRS Agar (BBL[®], USA); *Enterococcus* spp. on M-Enterococcus Agar (BBL[®], USA); Coliforms, including *E. coli* on Mac Conkey Agar (BBL[®], USA); *Clostridium* spp. – Clostridium Selective Agar (Merck, Germany). The plates were incubated anaerobically in Gas Pak[®] anaerobic system at 37°C for 96 h (TBC, *Bacteroides* and *Clostridium* spp.); at 37°C for 48 h (*Lactobacillus* and *Enterococcus* spp.), and aerobically at 37°C for 24 h (coliforms). *Cl. perfringens* enumeration was performed according to the Standard ISO 7937 (2004). This technique analyses all the toxin types of *Cl. perfringens*. The cultural medium used was Agar tryptose sulphite added with antibiotic d-cycloserine (Byrne et al., 2008). Agar plates were incubated at 37°C in Gas Pak[®] anaerobic system with carbon dioxide generating envelopes. *Cl. perfringens*

reduced sulphite to sulphide, and in the presence of iron, black colonies developed. These colonies were observed and counted.

After the incubation time had elapsed, the bacterial colonies were counted with an automatic colony counter. The total colony counts per g of undiluted caecal content from each medium were obtained as the weighed mean from two or three highest duplicate dilutions that showed growth. The bacterial counts were expressed as the average of five parallel replications of each sample in Log₁₀ colony forming units (CFU) per g caecal content (Log₁₀ CFU/g ± SEM).

Chemical analyses

After the caecal samples were taken for microbiological analyses the pH of the caecal digesta was measured immediately with a glass electrode pH meter. The chemical composition of the diet was analyzed according to standard methods of Association of Official Analytical Chemists (AOAC, 2000). Energy concentration of the diet was measured by adiabatic calorimeter.

Statistical analysis

All bacterial populations estimates were calculated and reported on a per g (wet weight) basis of caecal content. The mean and standard deviation of the bacterial counts were calculated using logarithmic values (Log₁₀ CFU/g). Statistical analysis of the results was performed by one-way analysis of variance (ANOVA). The results are quoted as means ± SEM.

Results and discussion

The caecal bacterial counts of rabbits weaned at 21 (W 21 group) and 35 day (W 35 group) is shown in Table 2. Results of the microbiological examination of the caecal contents indicated that at the days of weaning total bacterial counts (TBC) are 8.768 and 9.422 log₁₀ CFU/g, respectively. Rabbits weaned at 35 day had higher TBC per g of caecal content in comparison with rabbits weaned at 21 day (P<0.001). The TBC in the caecum of earlier and later weaned rabbits after weaning increased significantly (P<0.001). At the end of the experimental period (49 day) TBC in the caecal contents is 10.934 and 10.942 log₁₀ CFU/g, respectively. The differences between groups at the end of the trial, however, were never statistically significant. At this age the anaerobic bacterial flora in the

Table 2. Concentrations of different bacterial populations on caecal contents of rabbits (log₁₀ CFU/g)

Group/Day	Total Bacterial Count	<i>Bacteroides</i> spp.	<i>Clostridium</i> spp.	<i>Clostridium perfringens</i>	<i>Enterococcus</i> spp.	Coliforms	<i>Lactobacillus</i> spp.
W21 Group*							
21	8.768 ± 0.012 ^a	7.298 ± 0.015 ^a	3.354 ± 0.037 ^a	1.656 ± 0.101 ^a	6.494 ± 0.037 ^a	5.582 ± 0.067 ^a	<1
28	9.564 ± 0.022 ^b	8.250 ± 0.025 ^b	3.732 ± 0.033 ^b	2.202 ± 0.047 ^b	5.768 ± 0.221 ^b	4.472 ± 0.018 ^b	<1
35	10.836 ± 0.021 ^c	9.436 ± 0.033 ^c	3.964 ± 0.018 ^c	2.402 ± 0.024 ^c	4.674 ± 0.023 ^c	3.656 ± 0.022 ^c	<1
42	10.918 ± 0.008 ^d	9.756 ± 0.016 ^d	4.364 ± 0.043 ^d	2.526 ± 0.020 ^d	3.762 ± 0.053 ^d	3.194 ± 0.025 ^d	<1
49	10.934 ± 0.005 ^d	9.962 ± 0.011 ^{de}	5.446 ± 0.035 ^{de}	2.794 ± 0.071 ^{de}	3.242 ± 0.045 ^{de}	1.958 ± 0.037 ^{de}	<1
W35 Group**							
35	9.422 ± 0.018 ^e	8.164 ± 0.015 ^e	3.500 ± 0.063 ^e	1.644 ± 0.094 ^a	4.674 ± 0.023 ^c	4.314 ± 0.033 ^e	<1
42	10.624 ± 0.014 ^f	9.524 ± 0.017 ^f	4.218 ± 0.081 ^f	1.994 ± 0.032 ^f	3.462 ± 0.047 ^f	3.020 ± 0.018 ^f	<1
49	10.942 ± 0.013 ^d	9.856 ± 0.012 ^d	5.448 ± 0.046 ^{de}	2.408 ± 0.015 ^{dc}	3.086 ± 0.030 ^{dc}	1.865 ± 0.022 ^{dc}	<1

* Weaned at 21st day; ** Weaned at 35th day ; Significant differences in the same column and between groups: a, b, c, d, e, f, de, dc (P<0.001);

CE of the weaned rabbits develops and stabilizes, which is important for digestive health and productivity (Carabano et al., 2006). Several studies demonstrated also that strictly anaerobic bacteria are the main constituent of the CE in rabbits (Gouet and Fonty, 1979; Bónai et al., 2008; Monteils et al., 2008). Padilha et al. (1995) reported that the anaerobic bacterial flora in the caecum develops significantly from 3 weeks of age, as the feed intake of the young rabbits shifts from milk to dry feed.

Results of the microbiological examination of the caecal content indicate that the obligate anaerobic, Gram-negative, non-sporulating bacteria, particularly *Bacteroides* spp. constitute an important group of microorganisms in the rabbit caecum (Table 2). They were largely dominant compared with the other bacterial populations in the CE of young rabbits not only on the day of weaning, but later. During the experimental period the caecal population of *Bacteroides* spp. increased with advancing of age. The differences between the two groups on the day 35, 42 and 49 were statistically significant ($P < 0.001$). To the end of the trial the caecal number of *Bacteroides* spp. was significantly higher in early weaned rabbits ($P < 0.001$). Several studies also suggest that *Bacteroides* spp. is the main bacterial species in the rabbit's intestinal tract (Yanabe et al., 1999; Kovács et al., 2006). Some *Bacteroides* spp. play an important role on the immune response of the gut associated lymphoid tissue (Bry et al., 1996; Brubaker et al., 1999; Rhee et al., 2004; Mazmanian et al., 2005), and more knowledge on these species could be important to implement the intestinal health of rabbits.

Sporulating bacteria (*Clostridium* spp.) were about 1000 times less numerous than the *Bacteroides*, which are considered to belong to the sub-dominant flora. The total number of *Clostridium* spp. in the caecal content of early weaned rabbits increased from 3.354 (21 day) to 3.964 (35 day) and to 5.446 \log_{10} CFU/g (49 day) ($P < 0.001$). Similar results were obtained in the second experimental group where *Clostridium* count increased from 3.50 (35 day) to 5.448 \log_{10} CFU/g (49 day, $P < 0.001$). At the end of the trial (42 and 49 day) there were not significant differences in caecal concentrations of *Clostridium* spp. between groups (Table 2). Data from the current study show that *Cl. perfringens* was present in low variable amounts in all the caecal samples obtained from healthy animals. Caecal counts of *Cl. perfringens* at weaning (21 and 35 day) were very low (1.656 and 1.654 \log_{10} cfu/g, respectively) and not affected by weaning age. According to Maertens et al. (2006) rabbit milk contains antimicrobial compounds, such as medium chain fatty acids (caprylic and capric acids), that have been proved to have a significant bactericide effect (*in vitro*) against *Cl. perfringens* (Skrivanova et al., 2005).

The results from the current study indicate that after weaning caecal counts of *Cl. perfringens* were increased ($P < 0.001$), but rabbits weaned at 35 day had lower *Cl. perfringens* counts in comparison with rabbits weaned at 21 day ($P < 0.001$). To the end of the study earlier weaned rabbits had significantly higher caecal concentration of *Cl. perfringens* ($P < 0.01$) (Table 2). According to Chamorro et al. (2007), early substitution of solid feed for milk in early weaned rabbits decreased the digestibility of nutrients and increased the flow of nutrients reaching the ileum. An increase in the flow of ileal protein has been correlated with a higher frequency of detection of *Cl. perfringens* in the ileal contents. On the other hand, in early weaned rabbits intestinal digestion and absorption of starch is not as efficient as in adults and residual amounts of carbohydrate may reach the caecum to act as a substrate for bacterial fermentation. *Clostridium* spp. and other pathogenic bacteria can

proliferate in the caecum of young rabbits and can cause intestinal inflammation, enteritis, diarrhoea and rapid death due to the effects of enterotoxins (Harcourt-Brown, 2002). Marlier et al. (2006) and Szalo et al. (2007) suggest a correspondence between high *Cl. perfringens* counts ($> 2 \times 10^6$ CFU/g) and the incidence of epizootic rabbit enteropathy symptoms. According to Hatheway (1990), Songer (1996) and EFSA (2005) a high concentration of *Cl. perfringens* ($> 10^6$ CFU/g) in the caecal content is a predisposing factor for the release of toxins that causes clostridial enterotoxaemia.

Facultative anaerobic, Gram-positive bacteria belonging to the genera *Enterococcus* are an important part of the caecal microbial population of rabbits (Table 2). At the day of weaning the caecal count of *Enterococcus* spp. was higher in early weaned rabbits in comparison with rabbits weaned later (6.494 and 4.674 \log_{10} CFU/g), respectively. After weaning *Enterococcus* count in the caecal contents of groups A and B decreased considerably to the end of the trial to 3.242 and 3.086, respectively. The differences between the groups during the experimental period are statistically significant ($P < 0.01$).

Our study demonstrated that the coliforms count, including *E. coli* in the caecum of young rabbits, weaned at different age, is very low. The caecal number of coliforms was considerably high at weaning of the rabbits, then decreased linearly and stabilized on low level at day 49. The decrease in the level of coliforms between weaning and 49 days of age is in agreement with the results previously reported in healthy weaned rabbits (Gouet and Fonty, 1979; Padilha et al., 1999; Kovács et al., 2004a, 2008; Bónai et al., 2008). Our study demonstrated also significant differences between groups at 35 and 42 day when the caecal count of coliforms were higher in earlier weaned rabbits ($P < 0.01$). The higher number of coliforms in the caecum of earlier weaned rabbits to 42 day is considered to be a high risk from the animal health point of view (Bónai et al., 2008). Coliforms and especially *E. coli* are important opportunist pathogens and can be a major cause of enteritis and losses in rabbit farms. Under some circumstances pathogenic and toxigenic strains of the organism proliferate and cause diarrhea (Harcourt-Brown, 2002). Probably, early weaning (21 day) of rabbits makes them more conducive to coliform infections in comparison with later weaning (35 day).

Reports on the distribution of lactobacilli in the rabbit digestive tract are limited and those that are available have been inconsistent in results. Straw (1988) reported that cell population densities of lactobacilli in the rabbit gut were 10^6 per g of content. The results of the present experiment demonstrated the absence of *Lactobacillus* spp. in the rabbit caecal tract (Table 2). In most previous studies *Lactobacillus* spp. was also generally absent in the digestive tract of healthy rabbits (Gouet and Fonty, 1979; Penney et al., 1986; Straw (1988), Ducha et al., 1990; Harcourt-Brown, 2002). According to Pérez de Rozas et al. (2008) *Lactobacillus* spp. is not a habitual component of the microbiota of rabbits. Yu and Tsen (1993) also investigated the distribution of lactobacilli in the GIT of rabbits, but the results showed that they are present only in a few of the experimental rabbits and in small numbers. They studied the factors that might be responsible for such effects. These studies included investigation of the effect of gastric juice and bile salts on the viability of lactobacilli, assay of the survival rates of lactobacilli in the GIT of the rabbits and measurement of the adhesive capability of lactobacilli to the rabbit intestinal epithelial cells. Results showed that, although some lactobacilli were resistant to the rather low pH levels of rabbit gastric juice, lack of adhesive capability may prevent

them from colonizing in the intestinal tract.

There was considerable development of the caecum after weaning of the rabbits (Table 3). Age affected the empty caecum and the caecal contents. All parameters (CW and CC) significantly increased by the end of the trial (49 day) ($P < 0.001$). There were also significant differences between the two groups. Rabbits weaned at 35 day had significantly higher CW and CC at 35, 42 and 49 day in comparison with rabbits, weaned at 21 day ($P < 0.01$).

The pH of the caecal content fell linearly ($P < 0.01$) throughout the experiment (Table 3), from 6.78 on day 21 to 5.64 on day 49 (W 21 group) and from 6.78 on day 21 to 5.57 on day 49 (W 35 group). There are not significant differences between groups at the beginning (21 day) and at the end (49 day) of the trial. On the other

early weaned rabbits (W21 group), between days 21 and 28, and amounts in total to nearly 10%. The major part of these losses is due to diarrhea, whose etiology is multifactorial (Harcourt-Brown, 2002). On the other hand, weaning is a stressful period for young rabbits when healthy caecal microbiota is not yet established. The period around weaning is a very critical time for young rabbits as they are highly sensitive to multifactor digestive disorders (Kovacs et al., 2008). Up to 18-20 days, rabbits ingest only milk, while from this point onwards they begin to consume solid feed as well, initially in small quantities and subsequently, as their mother's milk yield diminishes in ever increasing quantities (Nizza et al., 2002). These results confirm the observations of Padilha et al. (1995) showing clearly that the change from milk to solid feed is much more

Table 3. Zootechnical and caecal weight traits of the rabbits weaned at different age

Trait	Age (day)				
	21	28	35	42	49
W 21 Group					
BW (g)	354 ^a (30)	456 ^b (22)	570 ^c (64)	954 ^d (102)	1240 ^e (100)
CW (g)	3.7 ^a (0.33)	6.9 ^b (0.36)	10.4 ^c (0.34)	16.4 ^d (0.36)	21.1 ^e (0.51)
CC (g)	6.5 ^a (0.40)	18.5 ^b (0.98)	24.6 ^c (1.54)	29.7 ^d (2.87)	37.7 ^e (2.55)
pH	6.78 ^a (0.13)	5.62 ^b (0.63)	5.61 ^b (0.36)	5.61 ^b (0.31)	5.64 ^b (0.19)
W 35 Group					
BW (g)	353 ^a (35)	860 ^c (16)	1206 ^f (52)	1410 ^g (155)	1780 ^h (134)
CW (g)	3.7 ^a (0.36)	7.1 ^b (0.23)	14.3 ^f (0.32)	19.7 ^g (0.43)	26.8 ^h (0.44)
CC (g)	6.4 ^a (0.25)	19.0 ^b (0.95)	29.2 ^f (1.25)	36.1 ^g (1.54)	45.4 ^h (2.74)
pH	6.78 ^a (0.13)	6.48 ^d (0.60)	6.54 ^d (0.40)	5.85 ^b (0.41)	5.57 ^b (0.19)

Data are mean (n=5) ± SEM (standard error of the mean).

Mean values in a same row with different superscripts were significantly different ($P < 0.001$)

Mean values in a same column with different superscripts were significantly different ($P < 0.01$)

hand, during the experimental period the early weaned rabbits (W 21 group) had significantly lower caecal pH only on day 28 and 35 ($P < 0.01$) in comparison with later weaned rabbits (W 35 group). The consistent decrease in pH between days 21 and 49 related to post weaning development of caecal microbial activity, fiber intake, and volatile fatty acid concentration in the caecum (Garcia et al., 2002; Bennegadi-Laurent et al., 2004).

The live body weight (BW) of the young rabbits had a typical evolution (Table 3). At weaning (21 and 35 day) the BW of the rabbits were respectively 354 ± 30 (W21 group) and 1206 ± 52 g (W35 group). These results were in agreement with McNitt and Moody (1988) and Scapinello et al. (1999), who reported a higher weaning weight when the pups consumed more milk during lactation. During the experimental period BW of the early weaned rabbits increased 1.6 times, between 21 and 35 day, and 1.47 times between days 35 and 49. Considerable differences between groups were also observed at the end of the experiment (49 day). Compared to the W21 group, rabbits in the W35 group, had a significantly higher BW (1240 ± 100 versus 1780 ± 134 g) ($P < 0.001$).

During the trial post-weaning mortality was observed only in

important than the weaning is from an exclusive milk intake to an exclusive solid feed intake. During this period, the GIT of rabbits is colonized by an abundant microflora, predominantly by strictly anaerobic bacteria (Marounek et al. 2000a). The fermentative activity of caecum begins to develop (Padilha et al., 1995, 1996; Piattoni et al., 1995) and the enzymatic digestive activities show important changes (Marounek et al., 1995). The microbial and fermentative activities in the CE of young rabbits play a key role in the prevention of digestive disorders (Carabaño et al., 2006). The pre-weaning milk/feed ratio affects the microbiological and biochemical characteristics of the caecal content and also post weaning mortality and performance of rabbits (Nizza et al., 2001).

Conclusion

Post-weaning developments and composition of the caecal microbiota in rabbits are influenced mainly by age and nutritional factors. The strictly anaerobic bacteria, particularly *Bacteroides* spp. were the main constituent of the caecal microbial ecosystem in the

weaned rabbits. *Clostridium* spp. was sub-dominant flora. Caecal counts of *Cl. perfringens* at weaning (21 and 35 day) were very low and not affected by weaning age. After weaning, to the end of the study, later weaned rabbits had lower *Cl. perfringens* counts in comparison with early weaned rabbits. *Enterococcus* spp. and coliforms were also an important part of the caecal microbial population of rabbits. The higher number of coliforms and especially *E. coli* in the caecum of earlier weaned rabbits is considered to be a high risk from the animal health point of view. *Lactobacillus* spp. is not a habitual component of the caecal microbiota of healthy rabbits weaned at different ages. Early weaning of rabbits did not cause detrimental changes of the caecal microbiota, but resulted in higher post weaning mortality and lower growth performance. Later weaning had positive effects related to post-natal development of the caecal microbial ecosystem, morbidity, mortality and live body weight of growing rabbits. However, further studies are necessary to acquire more information about this topic. The development and application of new molecular methods will provide a more extensive view of the caecal microbiota.

Acknowledgements

This study was financially supported by Faculty of Veterinary Medicine, Trakia University, 6000 Stara Zagora, Bulgaria (Research Project 11/08). The authors wish to express their full appreciation to Iliya Krachunov, PhD. from Feed Mill „Bonmix”, Lovech for the experimental diet preparation.

References

- Álvarez JL, Marguenda I, Garcia-Rebollar P, Carabano R, De Blas JC, Corujo A, and Garcia-Ruiz AI, 2007. Effects of type and level of fibre on digestive physiology and performance in reproducing and growing rabbits. *World Rabbit Science*, 15, 9-17.
- AOAC, 2000. Association of Official Analytical Chemists. Official Methods of Analysis, 17th Edition. AOAC, Arlington, VA, USA.
- Bennegadi-Laurent N, Gidenne T and Licois D, 2004. Nutritional and sanitary statuses alter post weaning development of caecal microbial activity in the rabbit. *Comparative Biochemistry and Physiology*, Part A, 139, 293-300.
- Bónai A, Szendro Zs, Maertens L, Matics Zs, Fébel H, Kametler L, Tornoyos G, Horn P, Kovács F, and Kovács M, 2008. Effect of inulin supplementation on caecal microflora and fermentation in rabbits. Proceedings of the 9th World Rabbit Congress, June 10-13, Verona, Italy, 555-559.
- Bry L, Falk PG, Midtvedt T and Gordon JI, 1996. A model of host-microbial interactions in an Open Mammalian Ecosystem. *Science*, 273, 1380-1383.
- Brubaker JO, Li Q, Tzianabos AO, Kasper DL and Finberg RW, 1999. Mitogenic activity of purified capsular polysaccharide A from *Bacteroides fragilis*: differential stimulatory effect on mouse and rat lymphocytes in vitro. *Journal of Immunology*, 162, 2235-2242.
- Byrne B, Scannell AGM, Lyng J and Bolton DJ, 2008. An evaluation of *Clostridium perfringens* media. *Food Control*, 19, 1091-1095.
- Carabano R, Badiola I, Licois D and Gidenne T, 2006. The digestive ecosystem and its control through nutritional or feeding strategies. In: *Recent Advances in Rabbit Sciences* (Eds. Maertens L and Coudert P), 211-227, ILVO, Merelbeke, Belgium.
- Carabano R, Badiola I, Chamorro S, Garcia J, Garcia-Ruiz AI, Garcia-Rebollar P, Gomes-Conde MS, Gutierrez I, Nicodemus N, Villamide MJ and De Blas JC, 2008. New trends in rabbit feeding: influence of nutrition on intestinal health. *Spanish Journal of Agricultural Research*, 6 (Special issue), 15-25.
- Chamorro S, Gómez-Conde MS, Pérez de Rozas AM, Badiola I, Carabaño R and De Blas JC, 2007. Effect on digestion and performance of dietary protein content and of increased substitution of lucerne hay with soya-bean protein concentrate in starter diets for young rabbits. *Animal*, 1, 651-659.
- Debray L, Gidenne T and Fortun-Lamothe L, 2001a. Evolution of intestinal enzymatic digestive capacity of rabbit around weaning. Proceedings of the 2nd Meeting COST 848, Workgroups 3 and 4, 29-30 June, Godollo, Hungary, pp. 37-38.
- Debray L, Gidenne T, Fortun-Lamothe L and Arveux P, 2001b. Efficacité digestive des lapereaux avant et après sevrage en fonction de la source énergétique du régime. Proceedings of the 9th Journées Recherche Cunicoles, Paris, France, pp. 191-194.
- Dojana N, Costache M and Dinischiotu A, 1998. The activity of some digestive enzymes in domestic rabbits before and after weaning. *Animal Science*, 66, 501-507.
- Ducha J, Lara C and Rodriguez AA, 1990. Correlation between airborne aerobic flora and intestinal flora in young rabbits bred in rabbitries. *Journal of Applied Rabbit Research*, 12, 228-230.
- EFSA, 2005. Opinion of the Scientific Panel on Biological Hazards on the request from the Commission related to *Clostridium* spp. in foodstuffs. *EFSA Journal*, 199, 1-65.
- Feugier A, Smit MN, Fortun-Lamothe L and Gidenne T, 2006. Fibre and protein requirements of early weaned rabbits and the interaction with weaning age: effects on digestive health and growth performance. *Animal Science*, 82, 493-500.
- Garcia J, Gidenne T, Falcao-E-Cunha L and De Blas C, 2002. Identification of the main factors that influence caecal fermentation traits in growing rabbits. *Animal Research*, 51, 165-173.
- Gidenne T and Fortun-Lamothe L, 2002. Feeding strategy for young rabbits around weaning: a review of digestive capacity and nutritional needs. *Animal Science*, 75, 169-184.
- Gidenne T, 2003. Fibers in rabbit feeding for digestive troubles prevention: respective role of low-digested and digestible fiber. *Livestock Production Science*, 81, 105-117.
- Gidenne T, Jehl N, Lapanouse A and Segura M, 2004. Inter-relationship of microbial activity, digestion and gut health in the rabbit: effect of substituting fibre by starch in diets having a high proportion of rapidly fermentable polysaccharides. *British Journal of Nutrition*, 92, 95-104.
- Gidenne T and Licois D, 2005. Effect of high fiber intake on the resistance of the growing rabbit to an experimental inoculation with an enteropathogenic strain of *Escherichia coli*. *Animal Science*, 80, 281-288.
- Gidenne T, Combes S, Licois D, Carabano R, Badiola I and Garcia J, 2008. Ecosystème caecal et nutrition du lapin: interactions avec la santé digestive. *INRA Production Animale* 21, 3, 239-250.
- Gomez-Conde MS, Garcia J, Chamorro S, Eiras P, Garcia Rebollar PG, Perez De Rozas A, Badiola I, De Blas C and Carabano R, 2007. Neutral detergent-soluble fiber improves gut barrier function in 25 d old weaned rabbits. *Journal of Animal Science*, 85, 3313-3321.
- Gouet Ph and Fonty G, 1979. Changes in the digestive microflora of holoxenic rabbits from birth until adulthood. *Annales de Biologie Animale, Biochimie, Biophysique*, 19, 553-566.
- Guide for the Care and Use of Laboratory Animals, 1996. Institute of Laboratory Animal Resources, Commission on Life

Sciences, National Research Council. National Academy Press, Washington, USA.

Gutierrez I, Espinosa A, Garcia J, Carabano R and De Blas JC, 2002. Effect of levels of starch, fiber, and lactose on digestion and growth performance of early weaned rabbits. *Journal of Animal Science*, 80, 1029-1037.

Hampson DJ, Pluske JR and Pethick DW, 2001. Dietary manipulation of enteric disease. In: *Digestive physiology of pigs* (Eds. Lindberg J.E., Ogle B.), 247-260, CABI Publishing.

Harcourt-Brown F, 2002. Textbook of Rabbit Medicine, 410 Butterworth, Heinemann, Oxford, UK.

Hatheway CL, 1990. Toxigenic clostridia. *Clinical Microbiology Review*, 3, 66-98.

Holdeman Moore, 1977. Anaerobe Laboratory Manual, 4th Edition, Virginia Polytechnic Institute and State University, Anaerobe Laboratory, Blacksburg, USA.

Hudson R, Shaal B, Martinez-Gomez M, and Distel H, 2000. Mother-young relations in the European rabbit: physiological and behavioral locks and keys. *World Rabbit Science*, 8, 85-90.

Kovacs M, Szendr Zs, Gyarmati T, Bencsne K, Donko T, Tornyo G, Lukacs H and Bota B, 2002. Effect of double nursing and early weaning on development of caecal microflora of the rabbit. *Magy Áo Lapja*, 124, 742-748 (Hg).

Kovacs M, Szendr Zs, Csutaras I, Bota B, Febel H, Kosa E, Bencs-Koll Z, Szakacs Á and Horn P, 2004. Some digestive-physiological parameters of early-weaned rabbits fed non-medicated diets. *Proceeding 8th World Rabbit Congress*, September 7-10, Puebla, Mexico, pp. 1097-1102.

Kovács M, Szendr Zs, Csutorás I, Bóta B, Fébel H, Kósa E, Bencs KZ and Balajca PK, 2004a. Some digestive physiological parameters of early weaned rabbits fed unmedicated diet. *Proceeding 16th Hungarian Conference on Rabbit Production*, Budapest, Hungary, pp. 33-38.

Kovács M, Szendr Zs, Milisits G, Bóta B, Bíró-Németh E, Radnai I, Pósa R, Bónai A, Kovács F and Horn P, 2006. Effect of nursing methods and faeces consumption on the development of the bacteroides, lactobacillus and coliform flora in the caecum of the newborn rabbits. *Reproduction Nutrition Development*, 46, 205-210.

Kovacs M, Milisits G, Szendr Z, Lukacs H, Bonai A, Posa R, Tornyo G, Kovacs F and Horn P, 2008. Effect of different weaning age (days 21, 28 and 35) on caecal microflora and fermentation in rabbits. *Proceeding - 9th World Rabbit Congress*, June 10-13, Verona, Italy, pp. 701-704.

Lavrencic A, 2007. The effect of rabbit age on *in vitro* caecal fermentation of starch, pectin, xylan, cellulose, compound feed and its fibre. *Animal*, 1, 241-248.

Maertens L and De Groote G, 1990. Feed intake of rabbit kit before weaning and attempts to increase it. *Journal of Applied Rabbit Research*, 13, 151-158.

Maertens L, Lebas F and Szendro Z, 2006. Rabbit milk: a review of quantity, quality and non-dietary affecting factors. *World Rabbit Science*, 14, 205-230.

Marlie D, Dewrée R, Lassence C, Licois D, Mainil J, Coudert P, Meulemans L, Ducatelle R and Vindevogel H, 2006. Infectious agents associated with epizootic rabbit enteropathy: isolation and attempts to reproduce the syndrome. *Veterinary Journal*, 172, 493-500.

Marounek M, Volvk SJ and Skrivanova V, 1995. Distribution of activity of hydrolytic enzymes in the digestive tract of rabbits. *British Journal of Nutrition*, 73, 463-469.

Marounek M, Brezina P and Baran M, 2000a. Fermentation of

carbohydrates and yield of microbial protein in mixed cultures of rabbit caecal microorganisms. *Archives of Animal Nutrition*, 53, 241-252.

Marounek M, Skrivanova V and Duskova D, 2000b. *In vitro* caecal fermentation of nitrogenous substrates in rabbits. *Journal of Agriculture Science Cambridge*, 135, 437-442.

Mazmanian SK, Liu CH, Tzianabos AO and Kasper DL, 2005. An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. *Cell*, 122, 107-118.

McNitt J and Moody GL, 1988. Milk intake and growth rates of suckling rabbits. *Journal of Applied Rabbit Research*, 11, 117-119.

Monteils V, Cauquil L, Combes S, Godon J and Gidenne T, 2008. Potential core species and satellite species in the bacterial community within the rabbit caecum. *FEMS Microbial Ecology*, 66, 620-629.

Nizza A, Di Meo C, Stanco G and Cutrignelli MI, 2001. Influence of solid feed intake and age at weaning on caecal characteristics and post-weaning performance. *World Rabbit Science*, 9, 149-153.

Nizza A, Stanco G, Dimeo C, Marongiu ML, Taranto S, Cutrignelli MI and Juliano L, 2002. Effect of pre-weaning solid feed and milk intake on caecal content characteristics and performance of rabbits around weaning. *Italian Journal of Animal Science*, 2, 95-101.

Padilha MTS, Licois D, Gidenne T, Carre B and Fonty G, 1995. Relationships between microflora and caecal fermentation in rabbits before and after weaning. *Reproduction Nutrition Development*, 35, 375-386.

Padilha MTS, Licois ISD and Coudert P, 1996. Frequency of the carriage and enumeration of *Escherichia coli* in caecal content of 15 to 49 day old rabbits. *Proceedings of the 6th World Rabbit Congress*, 3, Toulouse, France, pp. 99-102.

Padilha MTS, Licois D, Gidenne T and Carre B, 1999. Caecal microflora and fermentation pattern in exclusively milk-fed young rabbits. *Reproduction Nutrition Development*, 39, 223-230.

Parigi-Bini R and Xiccato G, 1998. Energy metabolism and requirements. In: *The nutrition of the rabbit*, (Eds. C. de Blas and J. Wiseman), CABI Publishing, Wallingford 103-132.

Pascual JJ, 2001. Recent advances on early weaning and nutrition around weaning. *Proceedings of the 2nd Meeting of workgroups 3 and 4, COST Action 848*, 29-30 June, 2001, Godollo, Hungary, 31-36.

Penney RL, Filk GE, Galask RP and Petzold CR, 1986. The microflora of the alimentary tract of rabbits in relation to pH, diet and cold. *Journal of Applied Rabbit Research*, 9, 152-156.

Pérez de Rozas AM, Rosell JM, Díaz JV, Carabaño R, García J, González J, Aloy N and Badiola I, 2008. Digestive microbiota studies in rabbits by REP-PCR method. *Proceeding 9th World Rabbit Congress*, June 10-13, Verona, Italy, pp. 1041-1044.

Piattoni F, Maertens L and Demeyer D, 1995. Age dependent variation of caecal contents composition of young rabbits. *Archives of Animal Nutrition*, 48, 347-355.

Pinheiro V, Gidenne T, Falcao E and Cunha L, 2001. Effect of age on bacterial fibrolytic activity of caecal flora of rabbit. *European Meeting COST 848, Workshop nutrition and pathology*, 29-30 June, Budapest, Hungary, pp. 50 (Abstr).

Rhee KJ, Sethupathi P, Driks A, Lanning DK and Knight KL, 2004. Role of commensal bacteria in development of gut associated lymphoid tissues and pre-immune antibody repertoire. *Journal of Immunology*, 172, 1118-1124.

Scapinello C, Gidenne T and Fortun-Lamothe L, 1999. Digestive capacity of the rabbit during the post-weaning period, according to the milk/solid feed intake pattern before weaning. *Reproduction*

Nutrition Development, 39, 423-432.

Schlolaut W, 1988. Present husbandry and management conditions and development trends in rabbit production. Proceedings - 4th World Rabbit Congress, 1, Budapest, Hungary, pp. 93-112.

Skrivanova E, Marounek M, Dlouha G and Kanja J, 2005. Susceptibility of *Clostridium perfringens* to c-2-c-18 fatty acids. Letters of Applied Microbiology, 41, 77-81.

Songer JG, 1996. Clostridial enteric diseases of domestic animals. Clinical Microbiology Review, 9, 216-234.

Standard ISO 7937, 2004. Microbiology of food and animal feeding stuffs-Horizontal method for the enumeration of *Clostridium perfringens* - Colony-count technique.

Stevens CE and Hume ID, 1995. Comparative Physiology of the Vertebrate Digestive System. 2nd Ed., Cambridge University Press, Cambridge, UK.

Straw TE, 1988. Bacteria of the rabbit gut and their role in the health of the rabbit. Journal of Applied Rabbit Research, 11, 142-50.

Szalo IM, Lassence C, Licois D, Coudert P, Poulipoullis A, Vindevogel H and Marlier D, 2007. Fractionation of the reference inoculum of epizootic rabbit enteropathy in discontinuous sucrose gradient identifies aetiological agents in high density fractions.

Veterinary Journal, 173, 652-657.

Xiccato G, Trocino A, Sartori A and Queaque PI, 2000. Early weaning rabbits: effect of age and diet on weaning and post weaning performance. Proceedings of the 7th World Rabbit Congress, Valencia, Spain, pp. 483-490.

Xiccato G, Trocino A, Sartori A and Queaque PI, 2001. Influence de l'âge, du sevrage précoce et de l'aliment sur le développement des organes digestifs caecales chez le jeune lapin. Proceedings of the 9th Journées Recherche Cunicoles, Paris, France, pp. 199-202.

Xiccato G, Trocino A, Sartori A and Queaque PI, 2003. Effect of weaning diet and weaning age on growth, body composition and caecal fermentation of young rabbits. Animal Science, 77, 101-111.

Yanabe M, Shibuya M, Gonda T, Asai H, Tanaka T, Narita T, Sudo K and Itoh K, 1999. Establishment of Specific Pathogen-Free Rabbit Colonies with Limited-Flora Rabbits Associated with Conventional Rabbit Flora, and Monitoring of Their Cecal Flora. Experimental Animals, 48, 101-106.

Yu B and Tsen HY, 1993. Lactobacillus cells in the rabbit digestive tract and the factors affecting their distribution. Journal of Applied Bacteriology, 75, 269-275.

Genetics and Breeding	
Selection of oil-bearing rose in Bulgaria – tendencies and perspective N. Kovatcheva	189
Combining ability of mutant maize line. I. Number of rows in the ear M. Ilchovska	193
Freezing of day 5 and 6 sheep and goat embryos of Greek breeds A. Pampukidou, M. Avdi, R. Ivanova T. Alifakiotis	196
Investigation on some seed characteristics among sunflower lines and hybrids M. Drumeva, N. Nenova, E. Penchev	199
Determination of coloured horses raised in Turkey O. Yilmaz, M. Ertugrul	203
Nutrition and Physiology	
Effects of different levels of dietary digestible amino acids on nitrogen retention and excretion in Topigs pig hybrids A. Ilchev, G. Ganchev	207
Development of the caecal microbiota in rabbits weaned at different age B. Bivolarski, G. Beev, S. A. Denev, E. Vachkova, T. Slavov	212
Consumption of dissolved oxygen in rainbow trout (<i>Oncorhynchus mykiss</i>) I. Sirakov, Y. Staykov, G. Djanovski	220
Effect of coconut oil on rumen and duodenal ammonia concentrations and some blood biochemistry parameters in yearling rams V. Radev, T. Slavov, E. Enev, I. Varlyakov	224
Pharmacokinetics of tiamulin and chlortetracycline after application of Tetramutin-premix in pigs D. Dimitrova V. Katsarov, D. Dimitrov, D. Tsoneva	229
Production Systems	
Research effect of application of herbicides raft 400 SC for growing of lavender D. Angelova, H. Lambev	235
Defining the critical kinematic parameters of rotary harrow with vertical axis of rotation D. Guglev	237
Development and experimental study of the maximum temperature potential of a solar thermal module for driving of an absorption air-conditioning machine K. Peychev, R. Georgiev	240
Histometrical investigation on the turkey broiler's third eyelid (Harderian) gland D. Dimitrov	246
Study of the tolerance of alfalfa varieties (<i>Medicago Sativa</i> L) to <i>Sitona</i> species (Coleoptera: <i>Curculionidae</i>) I. Nikolova, N. Georgieva	249
Productive performance and quality of essential oil from oil bearing rose (<i>Rosa damascena</i> Mill) for use of oxadiargyl D. Angelova	254
Study of the thermal efficiency of a solar thermal module at different mounting angles R. Georgiev, K. Peychev	257
Behavior of apple rootstock M9 produced by somatic organogenesis in stoolbed G. Dobrevska	261
Agriculture and Environment	
Effect of experimentally polluted water on the stomatal and structural characteristics on the leaves of two varieties of <i>Triticum aestivum</i> L. grown on different soil types K. Velichkova, D. Pavlov, D. Ninova	265
Ecological assessment of Cr (VI) concentrations in the surface waters of Stara Zagora Region used in agriculture N. Georgieva, Z. Yaneva, D. Dermendzhieva, V. Kotokova	269
Effect of shooting on the structure of population of golden jackal (<i>Canis aureus</i> L.) in Sarnena Sredna Gora mountain E. Raichev	276
Product Quality and Safety	
Chemical surface disinfection of funnel type fish egg incubators A. Atanasov, N. Rusenova, Y. Staykov, G. Nikolov, A. Pavlov, D. Stratev, E. Raichev	281
Fatty acid composition of common carp, rainbow trout and grey mullet fish species M. Stancheva, A. Merdzhanova	285

Instruction for authors

Preparation of papers

Papers shall be submitted at the editorial office typed on standard typing pages (A4, 30 lines per page, 62 characters per line). The editors recommend up to 15 pages for full research paper (including abstract references, tables, figures and other appendices)

The manuscript should be structured as follows: Title, Names of authors and affiliation address, Abstract, List of keywords, Introduction, Material and methods, Results, Discussion, Conclusion, Acknowledgements (if any), References, Tables, Figures.

The title needs to be as concise and informative about the nature of research. It should be written with small letter /bold, 14/ without any abbreviations.

Names and affiliation of authors

The names of the authors should be presented from the initials of first names followed by the family names. The complete address and name of the institution should be stated next. The affiliation of authors are designated by different signs. For the author who is going to be corresponding by the editorial board and readers, an E-mail address and telephone number should be presented as footnote on the first page. Corresponding author is indicated with *.

Abstract should be not more than 350 words. It should be clearly stated what new findings have been made in the course of research. Abbreviations and references to authors are inadmissible in the summary. It should be understandable without having read the paper and should be in one paragraph.

Keywords: Up to maximum of 5 keywords should be selected not repeating the title but giving the essence of study.

The introduction must answer the following questions: What is known and what is new on the studied issue? What necessitated the research problem, described in the paper? What is your hypothesis and goal?

Material and methods: The objects of research, organization of experiments, chemical analyses, statistical and other methods and conditions applied for the experiments should be described in detail. A criterion of sufficient information is to be

possible for others to repeat the experiment in order to verify results.

Results are presented in understandable tables and figures, accompanied by the statistical parameters needed for the evaluation. Data from tables and figures should not be repeated in the text.

Tables should be as simple and as few as possible. Each table should have its own explanatory title and to be typed on a separate page. They should be outside the main body of the text and an indication should be given where it should be inserted.

Figures should be sharp with good contrast and rendition. Graphic materials should be preferred. Photographs to be appropriate for printing. Illustrations are supplied in colour as an exception after special agreement with the editorial board and possible payment of extra costs. The figures are to be each in a single file and their location should be given within the text.

Discussion: The objective of this section is to indicate the scientific significance of the study. By comparing the results and conclusions of other scientists the contribution of the study for expanding or modifying existing knowledge is pointed out clearly and convincingly to the reader.

Conclusion: The most important consequences for the science and practice resulting from the conducted research should be summarized in a few sentences. The conclusions shouldn't be numbered and no new paragraphs be used. Contributions are the core of conclusions.

References:

In the text, references should be cited as follows: single author: Sandberg (2002); two authors: Andersson and Georges (2004); more than two authors: Andersson et al.(2003). When several references are cited simultaneously, they should be ranked by chronological order e.g.: (Sandberg, 2002; Andersson et al., 2003; Andersson and Georges, 2004). References are arranged alphabetically by the name of the first author. If an author is cited more than once, first his individual publications are given ranked by year, then come publications with one co-author, two co-authors, etc. The names of authors, article and journal titles in the Cyrillic or alphabet different from Latin, should be transliterated into Latin and article titles should be translated into English. The original language of articles and books translated into English is indicated in

parenthesis after the bibliographic reference (Bulgarian = Bg, Russian = Ru, Serbian = Sr, if in the Cyrillic, Mongolian = Mo, Greek = Gr, Georgian = Geor., Japanese = Ja, Chinese = Ch, Arabic = Ar, etc.)

The following order in the reference list is recommended:

Journal articles: Author(s) surname and initials, year. Title. Full title of the journal, volume, pages. Example:

Simm G, Lewis RM, Grundy B and Dingwall WS, 2002. Responses to selection for lean growth in sheep. *Animal Science*, 74, 39-50

Books: Author(s) surname and initials, year. Title. Edition, name of publisher, place of publication. Example: **Oldenbroek JK**, 1999. *Genebanks and the conservation of farm animal genetic resources*, Second edition. DLO Institute for Animal Science and Health, Netherlands.

Book chapter or conference proceedings: Author(s) surname and initials, year. Title. In: Title of the book or of the proceedings followed by the editor(s), volume, pages. Name of publisher, place of publication. Example:

Mauff G, Pulverer G, Operkuch W, Hummel K and Hidden C, 1995. C3-variants and diverse phenotypes of unconverted and converted C3. In: *Provides of the Biological Fluids* (ed. H. Peters), vol. 22, 143-165, Pergamon Press. Oxford, UK.

Todorov N and Mitev J, 1995. Effect of level of feeding during dry period, and body condition score on reproductive performance in dairy cows, IXth International Conference on Production Diseases in Farm Animals, Sept.11 – 14, Berlin, Germany, p. 302 (Abstr.).

Thesis:

Penkov D, 2008. Estimation of metabolic energy and true digestibility of amino acids of some feeds in experiments with muscovy duck (*Carina moschata*, L). Thesis for DSc. Agrarian University, Plovdiv, 314 pp.

The Editorial Board of the Journal is not responsible for incorrect quotes of reference sources and the relevant violations of copyrights.

AGRICULTURAL SCIENCE AND TECHNOLOGY

Volume 3, Number 3
September 2011



Journal web site:
www.uni-sz.bg/ascitech/index.html


Publisher:
www.alfamarket.biz